

## BBA Report

---

BBA 71341

### AMILORIDE-INDUCED STIMULATION OF $\text{HCO}_3^-$ REABSORPTION IN TURTLE BLADDER

GERHARD EHRENSPECK\*, JOHN DURHAM and WILLIAM A. BRODSKY\*\*

*Department of Physiology and Biophysics, Mount Sinai School of Medicine of the City University of New York, New York, N.Y. 10029 (U.S.A.)*

(Received November 21st, 1977)

#### Summary

Amiloride in the mucosal fluid (at concentrations of  $5 \cdot 10^{-6}$  M to  $10^{-4}$  M) reversibly stimulates the  $\text{HCO}_3^-$ -dependent moiety of the short-circuiting current ( $I_{sc}$ ) in ouabain-treated turtle bladders bathed by Na-free Ringer solutions with or without  $\text{Cl}^-$ .

This effect is uniquely different from the known inhibitory effect of this agent on  $\text{Na}^+$  transport. Thus, any comprehensive hypothesis on the action of amiloride over a wide dosage-response range should take into account its effect on  $\text{HCO}_3^-$  transport.

---

During a study of the possible adrenergic control of anion transport in turtle bladders bathed by Na-free Ringer solutions, the prior mucosal addition of amiloride ( $10^{-5}$  M to  $10^{-4}$  M) apparently prevented the previously reported [1, 2] stimulation of the anion-related short-circuiting current by the mucosal addition of norepinephrine ( $10^{-5}$  M to  $10^{-4}$  M). It was then found that the mucosal addition of amiloride alone produced a stimulation of this anion-related short-circuiting current across bladders in Na-free Ringer media. The present report deals with some characteristics of this amiloride-induced increase in anion reabsorption.

Procedures and methods for evaluating transepithelial potential ( $PD$ ), short-circuiting current ( $I_{sc}$ ), dc-resistance ( $R$ ), and  $^{36}\text{Cl}^-$  fluxes have been described [3, 4]. Bladders of *Pseudemys scripta* turtles were bathed on both surfaces by Na-free choline Ringer solutions plus  $10^{-4}$  M ouabain (Sigma) in

---

\*Present address: Department of Zoology and Microbiology and College of Osteopathic Medicine  
Ohio University, Athens, Ohio 45701, U.S.A.

\*\*To whom correspondence should be addressed.

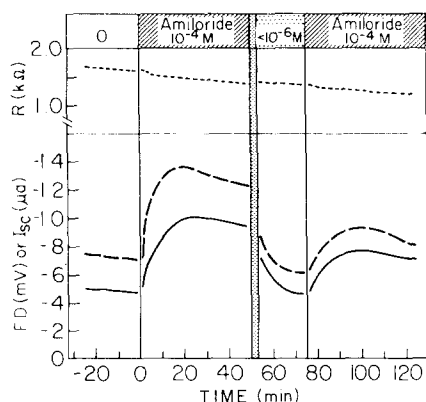


Fig. 1. Effect of  $10^{-4}$  M amiloride on  $PD$ ,  $I_{sc}$ , and  $R$  of ouabain-treated bladders in choline ( $Cl^- + HCO_3^-$ ) bathing system. Experimental conditions are described under Table I.  $R$  is depicted by dotted line in upper panel;  $PD$  is depicted by the dashed line and  $I_{sc}$  by the solid line in lower panel. Negative values of  $PD$  and  $I_{sc}$  denote transport of negative charges from M-to-S. Width of shaded vertical column denotes time required for reducing mucosal amiloride concentration by at least 6 sequential replacements of  $\frac{1}{2}$  the chamber volume (10 ml) with fresh bathing medium. This serial dilution procedure does not introduce the mechanical perturbations which in a complete replacement and rinsing of the mucosal compartment decrease transepithelial resistance. The magnitude of the second stimulation of  $PD$  and  $I_{sc}$  by  $10^{-4}$  M amiloride was about 60–70% of the first response.

the serosal fluid to inhibit any residual Na transport [5]. In some experiments Na-rich Ringer solutions plus ouabain were used.

Fig. 1 shows the effect of amiloride on a ouabain-treated bladder bathed on both surfaces by an identical Na-free, ( $Cl^- + HCO_3^-$ )-containing choline solution. The  $PD$  and  $I_{sc}$  increased within 0.5 min after the first addition of amiloride ( $10^{-4}$  M) to the mucosal fluid and reached a maximal value in about 20 min. Replacement of mucosal fluid by fresh Ringer solution restored the anion transport parameters to control levels. A second addition of amiloride to mucosal fluid resulted in a second increase in  $PD$  and  $I_{sc}$ .

Other results were the following: (i) amiloride in mucosal fluid stimulated the  $PD$  and  $I_{sc}$  in ouabain-treated bladders bathed by Na-rich ( $N = 3$ ) as well as the Na-free solutions; (ii) the threshold, half-maximal, and maximal responses to amiloride occurred at concentrations of  $10^{-5}$  M,  $3 \cdot 10^{-5}$  M, and  $10^{-4}$  M respectively; (iii) amiloride ( $10^{-3}$  M) in serosal fluid failed to stimulate the  $PD$  or  $I_{sc}$  ( $N = 2$ ); (iv) mucosal amiloride ( $10^{-4}$  M) failed to increase  $PD$  or  $I_{sc}$ , when these parameters were first stimulated by mucosal norepinephrine ( $10^{-4}$  M) ( $N = 3$ ).

Table I shows that in 13 bladders bathed by the ( $Cl^- + HCO_3^-$ ) solutions, amiloride induced significant increases in  $PD$  and  $I_{sc}$  and a small, but significant, decrease in transepithelial  $R$ . In three experiments on the mucosal to serosal (M-to-S) flux of  $^{36}Cl^-$ , the mean value of this flux was  $10.2 \pm 2.8 \mu A$  and remained unchanged ( $10.6 \pm 2.9 \mu A$ ) for 3 h after amiloride addition, whereas the  $I_{sc}$  was increased by  $65 \pm 9\%$  in 20 min.

Since amiloride did not change the M-to-S flow of  $Cl^-$ , it can be assumed that the amiloride-induced increment in the  $I_{sc}$  is due solely to an increase in the pumping of  $HCO_3^-$  from M-to-S. If this assumption is correct, amiloride should increase the  $I_{sc}$  (and  $PD$ ) in the presence of, but not in the

TABLE I

EFFECT OF AMILORIDE ON  $PD$ ,  $I_{sc}$ , AND  $R$  OF BLADDERS IN  $Cl^-$ -RICH,  $HCO_3^-$ -RICH BATHING SYSTEM

Mean values  $\pm$  S.E. ( $N = 13$ ) of  $PD$ ,  $I_{sc}$ , and  $R$  and of the percentage changes in these parameters at the time of maximal response after the mucosal addition of  $10^{-4}$  M amiloride. Sign convention: (M) electropositive to (S). Maximal response after  $22 \pm 5$  min. Area of exposed tissue,  $1.5 \text{ cm}^2$ . Composition of bathing fluid (mM): choline chloride, 21; choline bicarbonate, 20; choline sulfate, 30; KCl, 4;  $MgSO_4$ , 0.8;  $K_2 HPO_4$ , 0.61;  $KH_2 PO_4$ , 0.14;  $CaSO_4$ , 2; glucose, 11; sucrose, 20. Osmolality was 220 mosM/kg; final pH,  $7.5 \pm 0.1$ ; gassed with  $H_2O$ -saturated 98%  $O_2$  / 2%  $CO_2$ ; ouabain,  $10^{-4}$  M in (S).

Condition	$PD$ (mV)	$I_{sc}$ ( $\mu A$ )	$R$ ( $k\Omega$ )
Before amiloride	$18.2 \pm 3.8$	$11.0 \pm 2.0$	$1.6 \pm 0.1$
After amiloride	$28.7 \pm 5.5$	$19.7 \pm 3.0$	$1.4 \pm 0.1$
$\Delta(\%)^*$	$+76.8 \pm 12.2$	$+94.1 \pm 11.9$	$-9.9 \pm 2.0$
$P(\Delta=0)$	$< 0.001$	$< 0.001$	$< 0.001$

\*Note that values of  $\Delta(\%)$  are means  $\pm$  S.E. of  $N$  individual percentage changes in the designated electrical parameters after addition of amiloride. Statistical significance of  $\Delta$  was calculated by the Student's  $t$ -test.

TABLE II

EFFECT OF AMILORIDE ON  $PD$ ,  $I_{sc}$ , AND  $R$  OF BLADDERS BATHED BY  $HCO_3^-$ -RICH,  $Cl^-$ -FREE BATHING SYSTEM

Mean values  $\pm$  S.E. ( $N = 9$ ) of  $PD$ ,  $I_{sc}$ , and  $R$  and of the percentage changes in these parameters after mucosal addition of  $10^{-4}$  M amiloride. Composition of bathing fluid was similar to that described in Table I except that  $Cl^-$  was replaced by  $SO_4^{2-}$  without changing the choline and  $K^+$  concentrations. Osmolality was readjusted with sucrose. Maximal response after  $26 \pm 6$  min. Other experimental conditions and statistical definitions are given in Table I. Increases in values of  $PD$ ,  $I_{sc}$  were not significantly different from those in Table I ( $P > 0.05$  and  $P > 0.2$ , respectively)

Condition	$PD$ (mV)	$I_{sc}$ ( $\mu A$ )	$R$ ( $k\Omega$ )
Before amiloride	$13.7 \pm 2.8$	$8.3 \pm 2.4$	$2.5 \pm 0.5$
After amiloride	$29.0 \pm 6.0$	$16.3 \pm 4.6$	$2.4 \pm 0.5$
$\Delta(\%)$	$118 \pm 14$	$123 \pm 25$	$-4.7 \pm 4.1$
$P(\Delta=0)$	$< 0.001$	$< 0.001$	$> 0.2$

TABLE III

EFFECT OF AMILORIDE ON  $PD$ ,  $I_{sc}$ , AND  $R$  OF BLADDERS BATHED BY  $HCO_3^-$ -FREE,  $Cl^-$ -RICH MUCOSAL BATHING SYSTEM

Mean values  $\pm$  S.E. ( $N = 4$ ) of  $PD$ ,  $I_{sc}$ , and  $R$  and of the percentage changes in these parameters after mucosal addition of  $10^{-4}$  M amiloride. Composition of serosal fluid is described in Table I. Composition of mucosal fluid was similar to the serosal fluid except that  $HCO_3^-$  was replaced by  $SO_4^{2-}$ . Osmolality was readjusted with sucrose; final pH, 7.6; gassed with  $H_2O$ -saturated 100%  $O_2$ . Other experimental conditions, and statistical definitions are given in Table I.

Condition	$PD$ (mV)	$I_{sc}$ ( $\mu A$ )	$R$ ( $k\Omega$ )
Before amiloride	$10.1 \pm 2.1$	$8.2 \pm 0.8$	$1.3 \pm 0.3$
After amiloride	$9.6 \pm 2.4$	$7.0 \pm 0.9$	$1.4 \pm 0.3$
$\Delta(\%)$	$-7.4 \pm 11.4$	$-12.3 \pm 15.9$	$-9.7 \pm 5.9$
$P(\Delta=0)$	$> 0.5$	$> 0.4$	$> 0.1$

absence of  $\text{HCO}_3^-$  in mucosal fluid. This prediction was confirmed in the following experiments: First, with  $\text{HCO}_3^-$  but no  $\text{Cl}^-$  present in both mucosal and serosal fluid, the  $I_{\text{sc}}$  approximates the net reabsorption of  $\text{HCO}_3^-$  [4, 6]; under these bathing conditions, amiloride stimulated the  $I_{\text{sc}}$  (Table II). Secondly, with no exogenous  $\text{HCO}_3^-$  in the mucosal fluid ( $\text{Cl}^-$  present in mucosal, and  $\text{Cl}^-$  plus  $\text{HCO}_3^-$  present in serosal fluid), amiloride failed to stimulate the  $I_{\text{sc}}$  (Table III). Moreover, since under these bathing conditions the  $I_{\text{sc}}$  approximates the net  $\text{Cl}^-$  reabsorption [3], these data indicate a lack of effect by amiloride on  $\text{Cl}^-$  transport.

The data (Tables I–III) are completely accounted for by considering an amiloride-induced increase in the  $\text{HCO}_3^-$  moiety of  $I_{\text{sc}}$  and the hypothesis that the  $I_{\text{sc}}$  equals the algebraic sum of the conductive flows of  $\text{Cl}^-$  and  $\text{HCO}_3^-$  [4, 5, 7, 8]. An alternative hypothesis that the  $I_{\text{sc}}$  is due solely to the secretory pumping of protons and that all  $\text{Cl}^-$  reabsorption is non-conductive [9, 10] cannot account for the data. If the  $I_{\text{sc}}$  were generated by a proton secretion from cell to mucosal fluid, it should be stimulated by amiloride, whether or not  $\text{HCO}_3^-$  is present in the mucosal fluid. This is contrary to the facts (Table III).

Data from studies on the functional groups of amiloride (3,5-diamino-6-chloropyrazinoyl guanidine  $\cdot$  HCl) suggest that the guanidine group is required for the stimulation of  $\text{HCO}_3^-$  transport as well as for the inhibition of  $\text{Na}^+$  transport [11, 12, 13]. In 4 experiments, a guanidinium-free analogue (methyl-3, 5-diamino-6-chloropyrazinoate) failed to stimulate the  $PD$  or  $I_{\text{sc}}$  of bladders bathed by  $\text{HCO}_3^-$ -rich solutions in M and S.

The concentration of amiloride required to increase  $\text{HCO}_3^-$  reabsorption to half-maximal levels (i.e.,  $3 \cdot 10^{-5}$  M) was between 10 and 100 times more than that required to decrease the  $\text{Na}^+$  reabsorption by 50% in the turtle bladder [17], toad bladder [11], and mammalian kidney [16]. At similar concentrations (ca.  $10^{-4}$  M) in rat adipocyte preparations, amiloride has been found to decrease glucose transport and interfere with certain hormonally-induced decreases in carbohydrate, protein, and fat metabolism [14, 15]. Whereas these data show that higher concentrations of amiloride inhibit more than  $\text{Na}^+$  transport, the present evidence on  $\text{HCO}_3^-$  reabsorption is unique insofar as it reflects a stimulatory rather than an inhibitory effect of higher levels of amiloride.

The conclusions from these data on the turtle bladder are the following: (i) Amiloride stimulates the active reabsorption of  $\text{HCO}_3^-$  ions and this effect is independent of the inhibition of  $\text{Na}^+$  transport by this drug. Consequently, amiloride is not a modifier of cation transport [11, 16] alone, as has been conventionally assumed, (ii) To account for all of the observations, one is forced to invoke some interaction(s) between amiloride and the apical membrane other than an increase in the positive surface charge density. Although such an increase in surface charge could account for the direction of change in  $PD$ , the stimulation of  $\text{HCO}_3^-$  transport (Table II), and the inhibition of  $\text{Na}^+$  transport [17], it cannot account for the lack of effect on the M-to-S flux of  $\text{Cl}^-$ , whether this  $\text{Cl}^-$  flux is driven by an electrogenic pump [3, 8, 18] or an electroneutral anion exchanger [9, 10].

The similarity of the amiloride and norepinephrine-induced [1, 2] sti-

mutations of  $\text{HCO}_3^-$ -transport in the turtle bladder would suggest that their actions involve common mechanisms in the apical membrane [2].

### Acknowledgement

Amiloride and its analogue was kindly supplied by Dr. E.J. Cragoe of Merck Sharp and Dohme, West Point, Pa. This study was supported by grants from the NIH (1 R01 AM-16928-03) and NSF (PCM76-02344). Dr. G. Ehrenspeck was supported by U.S.P.H.S. Training grant (1 T32-EY07014-02).

The authors wish to show their appreciation to Mrs. Cristina Matons and Mrs. S. Ehrenspeck for their expert technical assistance.

### References

- 1 Brodsky, W.A., Schilb, T.P. and Parkes, J.L. (1976) in *Gastric Hydrogen Ion Secretion* (Kasbekar, Sachs, and Rehm, eds.), Vol. 3, Chap. 17 pp. 404—432, Marcel Dekker Co., New York
- 2 Brodsky, W.A. and Ehrenspeck, G. (1977) in *Membrane Toxicity* (Miller, M.W. and Shamoo, A.E., eds.), pp. 41—66, Plenum Press, New York
- 3 Gonzalez, C.F., Shamoo, Y.E. and Brodsky, W.A. (1967) *Am. J. Physiol.* 212, 641—650
- 4 Gonzalez, C.F. (1969) *Biochim. Biophys. Acta* 193, 146—158
- 5 Solinger, R.S., Gonzalez, C.F., Shamoo, Y.E., Wyssbrod, H.R. and Brodsky, W.A. (1968) *Am. J. Physiol.* 215, 249—260
- 6 Gonzalez, C.F. and Schilb, T.P. (1969) *Biochim. Biophys. Acta* 193, 419—429
- 7 Gonzalez, C.F., Shamoo, Y.E. and Brodsky, W.A. (1969) *Biochim. Biophys. Acta* 193, 403—418
- 8 Gonzalez, C.F., Shamoo, Y.E., Wyssbrod, H.R., Solinger, R.E. and Brodsky, W.A. (1967) *Am. J. Physiol.* 213, 333—340
- 9 Schwartz, J.H. (1976) *Am. J. Physiol.* 231, 565—572
- 10 Leslie, B.R., Schwartz, J.H. and Steinmetz, P.R. (1973) *J. Physiol.* 225, 610—617
- 11 Bentley, P.J. (1968) *J. Physiol.* 195, 317—330
- 12 Benos, D.W., Simon, S.A., Mandel, L.J. and Cala, P.M. (1976) *J. Gen. Physiol.*, 68, 43—63
- 13 Cuthbert, A.W. (1976) *Mol. Pharmacol.* 12, 945—957
- 14 Bruchhausen, F.V., Kaiser, I. and Herken, H. (1969) *N.S. Arch. Pharmak. Exp. Pathol.* 262, 139—151
- 15 Bruchhausen, F.V. and Streubel, J. (1969) *N.S. Arch. Pharmak. Exp. Pathol.* 262, 251—263
- 16 Baer, J.E., Jones, C.B., Spitzer, S.A. and Russo, H.F. (1967) *J. Pharmacol. Exp. Therap.* 157, 472—485
- 17 Wilczewski, T. and Brodsky, W.A. (1975) *Am. J. Physiol.* 228, 781—790
- 18 Brodsky, W.A. and Schilb, T.P. (1966) *Am. J. Physiol.* 210, 987—996